

## JERICO-RI: Progress toward an automated detection of phytoplankton in European coastal areas



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## Automated platform for the observation of phytoplankton diversity in relation to ecosystem services

A large set of instruments to qualify and expertise: SYKE, SMHI, HZG, RWS, VLIZ, CEFAS, CNRS (LOG, BOREA, OSU-V, MIO) & Ifremer (LER/BL)



- Pulse-shape recording flow cytometer (Cytosense)
- Imaging in-flow (imaging in-flow Cytobot)
- FlowCAM
- FastCAM
- Spectral fluorometer (AOA, Fluoroprobe, Multiexciter)
- Absorption meter (PSI-CAM)
- Pulse Amplitude Modulated Fluorometers (PAM)
- Fast Repetition Rate Fluorometer (FRRF)
- Underwater Vision Profiler (UVP5)
- (Semi-)Automated data analysis

#### Outcomes:

A set of **recommendations** of the most suitable and relevant **combination of methods** according to the environment considered, their limits and ways of implementing them as **complementary sensors in combined platforms**. A JERICO-NEXT deliverable to come by one year.

Phytoplankton groups discriminated by automated pulse-shape recording flow cytometry (Cytobuoy b.v.)

## nation of methods according to the



Chlorophyll a equivalent of the multispectral fluorometry (fluoroprobe bbe Moldaenke): discrimination of 4 pigmentary groups (Haptophytes, brown algae, phycocyanin and Phycoerythrin) containing micro-algae during a Phaeocystis globosa bloom

# EEC (UK) \*\*\* Characterisms\*\*\* \*\*\* Printingstand\*\*\* \*\*\* Printingstand\*\* \*\*\* Printingstand\*\*\* \*\*\* Printingstand\*\* \*\*\* Printingstand\*\*\* \*\*\* Printingstand\*\* Printingstand\*\* \*\*\* Printingstand\*\* \*\*\* Pri

JERICO-NEXT spring Channel-North Sea cruises (April-May 2017)

### Up to 13 groups discriminated

<u>Waters under brackish influence</u>: Dominated by nanophytoplankton (and microphytoplankton).

Offshore and other coastal waters:
Dominated by picoeukaryotes, Synechococcus-like (and
NanoSWS coccolithophores-like)



**EEC-SNS JERICO-NEXT 2017 cruises** 

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Same trend between the total fluorescence from the ferrybox and the total red fluorescence from the flow cytometer (FLR-cytosense) during the survey.

Distribution of the number of particles.ml-1 according to the phytoplankton functional types



- $\checkmark$  More than 90 % of the particles = phytoplankton,  $\sim$  93% of the total fluorescence.
- ✓ No relationship between salinity, temperature and the number of particles per functional types: Synechococcus, picophytoplankton, nanoplankton and microphytoplankton.
- FerryBox: Fluorescence (Fluores), Quantum ✓ Difference in the distribution of the phytoplankton functional types with a higher yield (QY), Salinity (SAL), Temperature (SST) density of nanoplankton in the north of the area.

Intercomparison exercise: 3 pulse shape recording automated low cytometers (fcm) (VLIZ, RWS and CNRS-LOC) during the JERICO-NEXT – LifeWatch spring cruise 2017 (8–12 May 2017).led by VLIZ

- ✓ Similar trends in measured total cell/colony abundance per ml.
- ✓ Automatedfcm: high potential for large scale data collection at high temporal and spatial resolution.
- ✓ Harmonizing the sampling protocol will increase data comparison and greatly improve the insights into phytoplankton community dynamics.



